



# TWIST1 Promotes Invasion Through Mesenchymal Change in Human Glioblastoma

## Citation

Mikheeva, Svetlana A., Andrei M. Mikheev, Audrey Petit, Richard Beyer, Robert G. Oxford, Leila Khorasani, John-Patrick Maxwell, et. al. 2010. TWIST1 promotes invasion through mesenchymal change in human glioblastoma. *Molecular Cancer* 9: 194.

## Published Version

doi://10.1186/1476-4598-9-194

## Permanent link

<http://nrs.harvard.edu/urn-3:HUL.InstRepos:5335447>

## Terms of Use

This article was downloaded from Harvard University's DASH repository, and is made available under the terms and conditions applicable to Other Posted Material, as set forth at <http://nrs.harvard.edu/urn-3:HUL.InstRepos:dash.current.terms-of-use#LAA>

## Share Your Story

The Harvard community has made this article openly available.  
Please share how this access benefits you. [Submit a story](#).

[Accessibility](#)

RESEARCH

Open Access

# TWIST1 promotes invasion through mesenchymal change in human glioblastoma

Svetlana A Mikheeva<sup>1,3†</sup>, Andrei M Mikheev<sup>1,3,7†</sup>, Audrey Petit<sup>1,3</sup>, Richard Beyer<sup>2</sup>, Robert G Oxford<sup>1</sup>, Leila Khorasani<sup>1</sup>, John-Patrick Maxwell<sup>1</sup>, Carlotta A Glackin<sup>4</sup>, Hiroaki Wakimoto<sup>5</sup>, Inés González-Herrero<sup>6</sup>, Isidro Sánchez-García<sup>6</sup>, John R Silber<sup>1</sup>, Philip J Horner<sup>1,3</sup>, Robert C Rostomily<sup>1,3\*</sup>

## Abstract

**Background:** Tumor cell invasion into adjacent normal brain is a mesenchymal feature of GBM and a major factor contributing to their dismal outcomes. Therefore, better understandings of mechanisms that promote mesenchymal change in GBM are of great clinical importance to address invasion. We previously showed that the bHLH transcription factor TWIST1 which orchestrates carcinoma metastasis through an epithelial mesenchymal transition (EMT) is upregulated in GBM and promotes invasion of the SF767 GBM cell line *in vitro*.

**Results:** To further define TWIST1 functions in GBM we tested the impact of TWIST1 over-expression on invasion *in vivo* and its impact on gene expression. We found that TWIST1 significantly increased SNB19 and T98G cell line invasion in orthotopic xenotransplants and increased expression of genes in functional categories associated with adhesion, extracellular matrix proteins, cell motility and locomotion, cell migration and actin cytoskeleton organization. Consistent with this TWIST1 reduced cell aggregation, promoted actin cytoskeletal re-organization and enhanced migration and adhesion to fibronectin substrates. Individual genes upregulated by TWIST1 known to promote EMT and/or GBM invasion included SNAI2, MMP2, HGF, FAP and FN1. Distinct from carcinoma EMT, TWIST1 did not generate an E- to N-cadherin “switch” in GBM cell lines. The clinical relevance of putative TWIST target genes SNAI2 and fibroblast activation protein alpha (FAP) identified *in vitro* was confirmed by their highly correlated expression with TWIST1 in 39 human tumors. The potential therapeutic importance of inhibiting TWIST1 was also shown through a decrease in cell invasion *in vitro* and growth of GBM stem cells.

**Conclusions:** Together these studies demonstrated that TWIST1 enhances GBM invasion in concert with mesenchymal change not involving the canonical cadherin switch of carcinoma EMT. Given the recent recognition that mesenchymal change in GBMs is associated with increased malignancy, these findings support the potential therapeutic importance of strategies to subvert TWIST1-mediated mesenchymal change.

## Background

Invasion is arguably the feature of human glioblastoma (GBM) most responsible for their dismal outcomes with average survival less than 1 year. Diffuse tumor invasion into adjacent brain restricts curative resection and limits effective delivery of chemotherapy and radiation. In addition, migratory GBM cells can activate mechanisms that increase resistance to these therapies further compounding efforts to eradicate them. Despite the importance of glioma invasion, little is known about how this

complex phenotype is regulated in gliomas, a prerequisite to development of effective anti-invasion therapies. By contrast, the process by which human epithelial cancers, or carcinomas, acquire an invasive phenotype has been more extensively characterized at both the cellular and molecular levels.

Carcinoma invasion and metastasis are driven by a process termed epithelial to mesenchymal transition (EMT) (For review see [1]). Mesenchymal transitions lead to acquired potential for cell migration, changes in cytoskeletal organization, reduced cellular adhesion and changes in expression of transcription factors. Among the transcription factors that play fundamental roles in regulating these changes is the basic helix-loop-helix

\* Correspondence: [rosto@u.washington.edu](mailto:rosto@u.washington.edu)

† Contributed equally

<sup>1</sup>Department of Neurological Surgery, University of Washington School of Medicine, Seattle WA, 98195, USA

protein TWIST1. TWIST1 activates EMT in the context of embryonic morphogenesis [2], tissue fibrosis [3,4] and cancer metastasis [5-7]. A central feature of TWIST1-mediated EMT is the repression of the epithelial marker E-cadherin, and activation of the mesenchymal marker N-cadherin [5-7], a hallmark feature of carcinoma EMT termed the “cadherin switch”. The recent recognition of mesenchymal change in glioblastoma [8-10] and its association with more aggressive clinical phenotypes [8,9] suggests that mechanisms that promote EMT in carcinoma may be of great clinical relevance in GBM.

We previously reported that TWIST1 is up-regulated in malignant gliomas and promotes glioma cell invasion of the SF767 glioma cell line *in vitro* [11]. However, the role of TWIST1 in promoting glioma invasion has not been investigated in the context of the brain microenvironment or as a mediator of mesenchymal change as occurs in carcinomas. In addition, the identification and clinical relevance of putative TWIST1 target genes in GBMs is not known. In this study we report that TWIST1 promoted GBM invasion through activation of mesenchymal molecular and cellular changes. This effect was not dependent on a “cadherin switch” indicating that TWIST1 promotes invasion through mesenchymal changes distinct from those associated with carcinoma EMT. The highly correlated expression of TWIST1 and mesenchymal target genes SNAI2 and FAP in human gliomas supported the clinical relevance of TWIST1 mesenchymal change. Together these results demonstrated an important role of TWIST1 in glioma invasion through activation of mesenchymal change and suggest its potential as a therapeutic target.

## Results

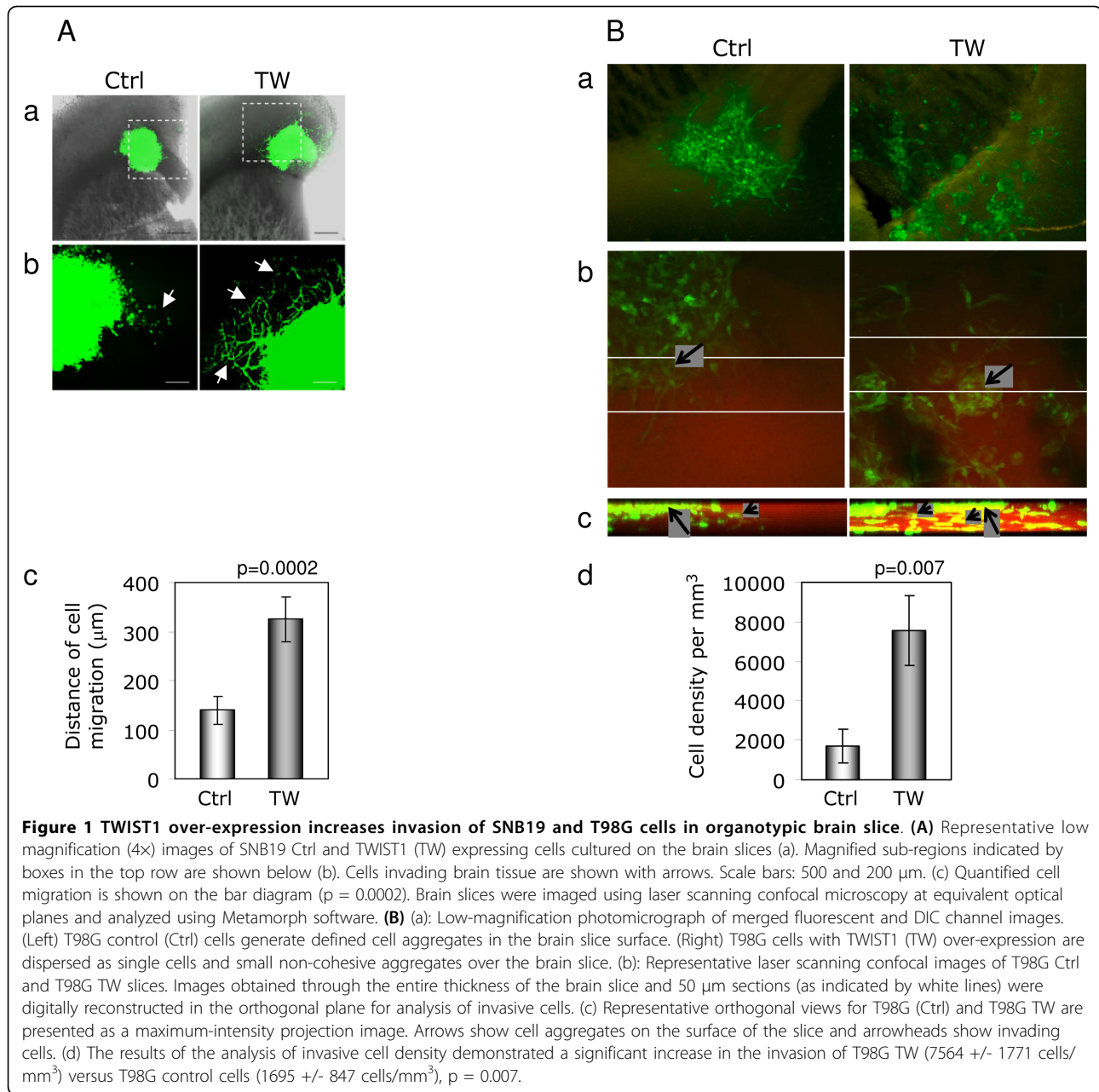
### **TWIST1 regulates invasion of multiple GBM cell lines *in vitro***

To extend our previous observation that TWIST1 enhanced invasion of the SF767 GBM cell line *in vitro* [11], we studied the effects of TWIST1 over-expression on invasion of SNB19 and T98G GBM cells using matrigel transwell assays. Compared with controls, over-expression of TWIST1 in SNB19 cells (SNB19 TW) and T98G cells (T98G TW) resulted in an increase of invasion of 68% and 80%, respectively (Additional file 1). The pro-invasive function of TWIST1 was also confirmed in a well-characterized primary GBM stem cell line (GBM4) [12] where a five-fold increase in TWIST1 expression resulted in a 140% increase in invasion *in vitro* (Additional file 2). Together, these results solidly established the physiologic importance of TWIST1 for GBM invasion by demonstrating its uniformly pro-invasive function in multiple GBM cell lines *in vitro*.

### **TWIST1 promotes GBM cell invasion in brain slice cultures and intact brain *in vivo***

To demonstrate pro-invasive TWIST1 function in more clinically relevant contexts, we characterized the growth patterns of SNB19 TW, T98G TW and corresponding control cells in an *ex vivo* model using organotypic brain slice cultures and *in vivo* using an orthotopic xenotransplant model. In *ex vivo* analyses, both SNB19 TW and T98G TW cells were significantly more invasive than control cells (Figure 1). Confocal microscopy confirmed that cells invaded into the brain slice rather than simply migrating along the surface. For SNB19, TWIST1 over-expression resulted in significantly increased distances of invasion compared with control cells when measured as orthogonal distances of cell migration from the border of the cohesive cell aggregate ( $p = 0.0002$ ) (Figure 1A). T98G control cells placed on the brain slice formed a cohesive cell aggregate but T98G TW cells did not form a central cohesive core in 7 of 8 slices and diffused from the implant site into the brain slice as small cell clumps (Figure 1B). Therefore, for T98G we analyzed invasive cell density through the entire thickness of the brain slice using orthogonal reconstruction of confocal optical slices. Results shown in Figure 1B demonstrate that invading density of T98G TW cells is 4 fold higher compared to invading cell density of control cells.

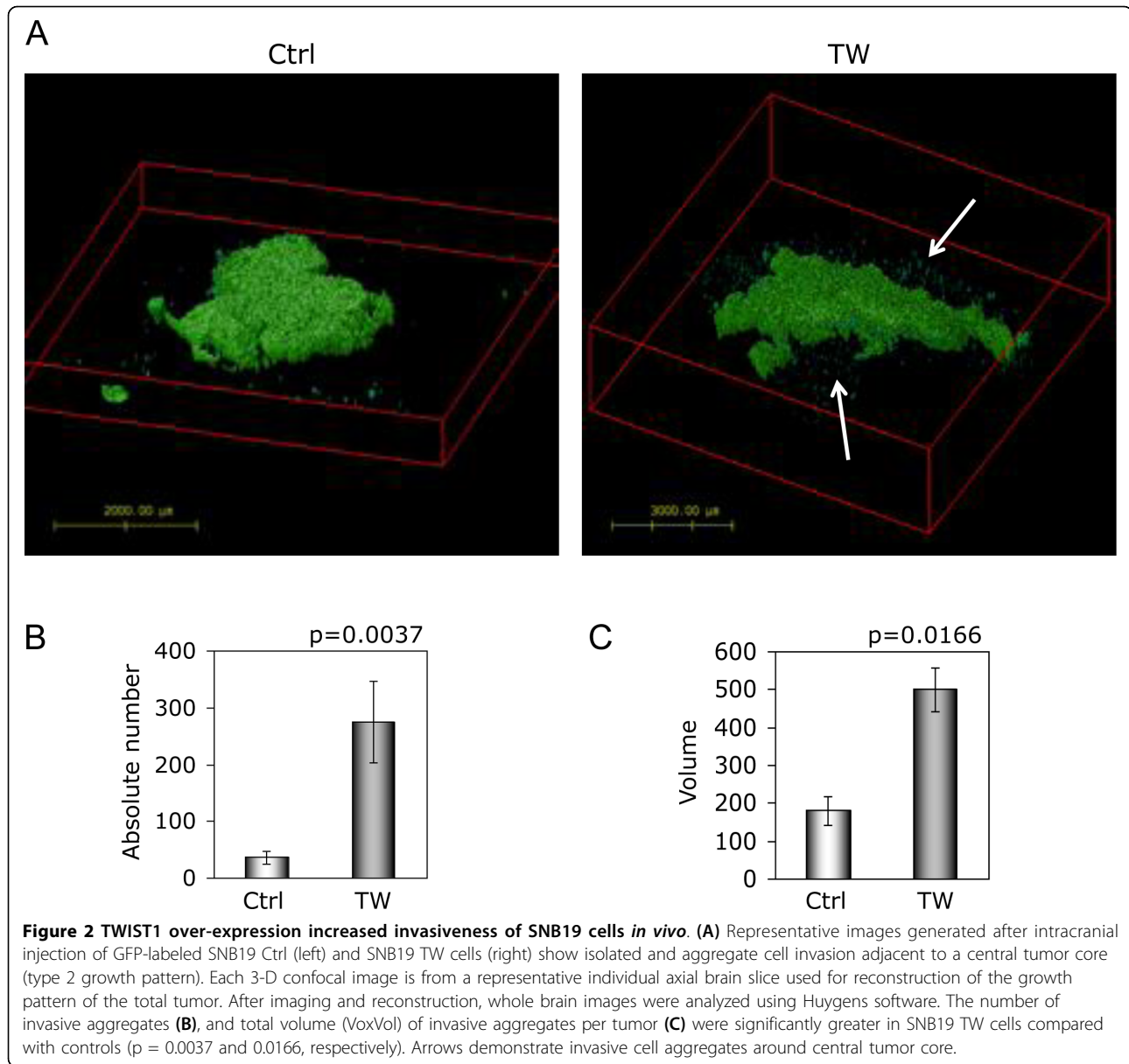
To determine whether TWIST1 over-expression increased invasion in the intact brain, we implanted SNB19 TW and T98G TW and corresponding control cells stably expressing GFP protein into the caudate nucleus of immuno-compromised adult mice (SNB19 TW ( $n = 6$ ), control ( $n = 4$ ); T98G TW and control ( $n = 3$  each)). All animals were sacrificed when neurologic morbidity was apparent in the first animal (day 17 for SNB19 and day 90 for T98G). Whole brain fluorescent imaging by laser scanning confocal microscopy followed by computer-aided digital image reconstruction provided a comprehensive global comparison of the tumor growth patterns throughout the entire brain of each animal. SNB19 TW and control tumors all met criteria for Type 2 pattern of invasion with a central core surrounded by individual invasive cells and cell aggregates. Therefore, to determine differences in invasiveness between SNB19 TW and control tumors we used Huygens image analysis software to quantify the number of fluorescent particles detected as discrete objects separate from the tumor core (indicating migratory tumor cells or clumps), the composite volume of these particles for each tumor and finally the core tumor volumes. A representative reconstructed tumor slice volume showing cores and invasive particles is shown in Figure 2A. This analysis demonstrated a significant increase in the number of discrete aggregates separate from the tumor



cores and the volume of the invasive particles in tumors with TWIST1 over-expression compared to control tumors ( $p = 0.0037$  and  $p = 0.0166$ , respectively) (Figure 2B, C) and a trend towards larger mean tumor core volume in SNB19 TW tumors ( $p = 0.11$ ). All T98G control tumors grew as localized expansile masses (Type 1 growth pattern) while all T98G TW tumors generated markedly invasive tumors that disseminated diffusely throughout the brain (Type 3 growth pattern) (Figures 3 and 4). Individual optical sections from corresponding areas of T98G control and TWIST1 tumors (Figure 3) and a brightest point projection image (BPI) (Figure 4)

which sums the individual signals through the entire Z-stack onto a 2-dimensional image, demonstrated the differences in growth patterns. The growth pattern for all tumors generated from T98G TW cells appeared to coincide with white matter tracts and crossed midline (Figure 3), both recognized primary routes of human GBM tumor cell invasion. Taken together these results provided critical confirmation of the pro-invasive function of TWIST1 in GBM cells in the context of the brain microenvironment and demonstrated cell-type-specific variability in TWIST1-mediated patterns of invasion.

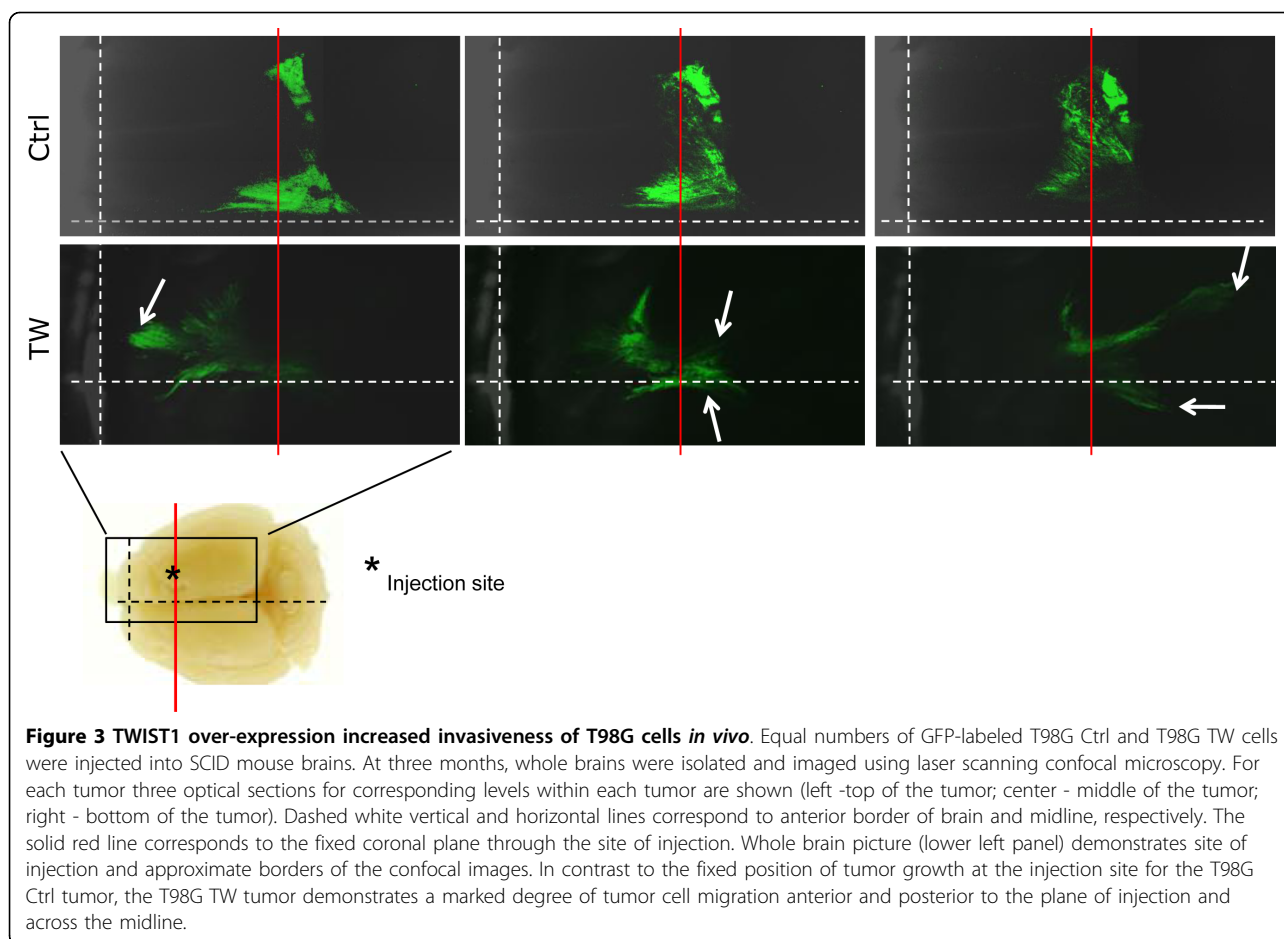




#### TWIST1 activates cell type-specific gene expression profiles consistent with invasion

To characterize the molecular basis of TWIST1 expression and associated pro-invasive phenotypes we compared expression profiles of SNB19 and T98G cells with TWIST1 over-expression and corresponding control cells using Affymetrix gene expression arrays. Overall, TWIST1 over-expression led to differential regulation of 1924 genes in SNB19 and 1525 genes in T98G cell lines (1.5 fold,  $p < 0.05$ ). Among these, 189 common genes were differentially regulated by TWIST1 over-expression in both cell lines (Pearson's correlation 0.557;  $p < 0.05$ ; Additional file 3). Using GO analysis we determined biological process gene categories that were significantly

altered by TWIST1 over-expression (FDR = 0.1). In both cell lines TWIST1 over-expression resulted in consistent and significant over-representation of genes in GO biological process categories including cell adhesion, extracellular matrix, cell motility and locomotion, cell migration and actin cytoskeleton organization (Figure 5). Of note, these categories are fundamental biological processes that are component features collectively required to achieve cell invasion [1,13]. GO biological process categories unique to SNB19 included enzyme-linked receptors and transcription while T98G cell type-specific categories included development, morphogenesis and intracellular signaling cascade. Of interest, a significant over-representation of genes in nervous system



**Figure 3 TWIST1 over-expression increased invasiveness of T98G cells *in vivo*.** Equal numbers of GFP-labeled T98G Ctrl and T98G TW cells were injected into SCID mouse brains. At three months, whole brains were isolated and imaged using laser scanning confocal microscopy. For each tumor three optical sections for corresponding levels within each tumor are shown (left -top of the tumor; center - middle of the tumor; right - bottom of the tumor). Dashed white vertical and horizontal lines correspond to anterior border of brain and midline, respectively. The solid red line corresponds to the fixed coronal plane through the site of injection. Whole brain picture (lower left panel) demonstrates site of injection and approximate borders of the confocal images. In contrast to the fixed position of tumor growth at the injection site for the T98G Ctrl tumor, the T98G TW tumor demonstrates a marked degree of tumor cell migration anterior and posterior to the plane of injection and across the midline.

development, a daughter category with the parent category of development, was also noted in T98G TW cells. The specific genes regulated by TWIST1 in these cell lines are discussed in more detail below. While this analysis demonstrated that cell type-specific changes in functional gene expression occur in GBM cells over-expressing TWIST1, we found that TWIST1 most consistently activated common molecular motifs related to cell invasion in both GBM cell lines.

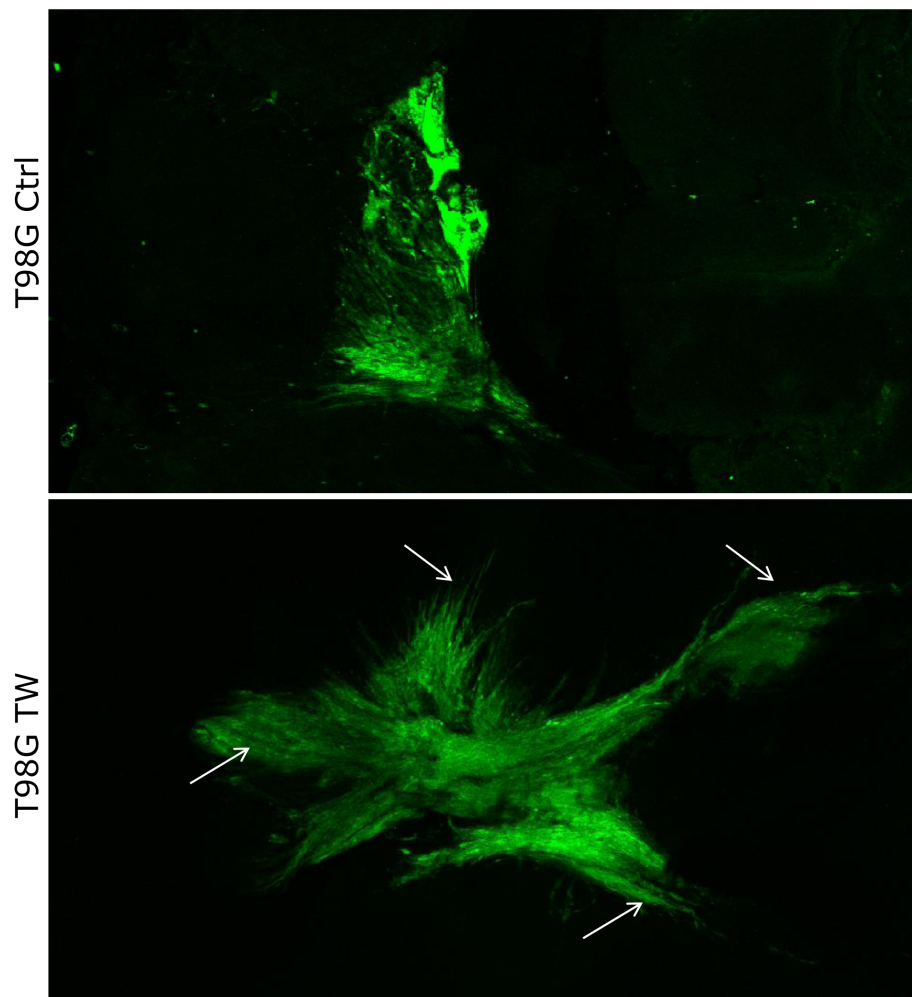
#### **TWIST1 mediated alteration of cell-cell adhesion, cell-substrate interactions, migration, and actin cytoskeleton in glioma cells**

To determine whether the GO analysis was consistent with TWIST1-mediated cellular phenotypes we performed functional assays in SNB19 TW cells for changes in cell-cell adhesion, cell-substrate interactions, migration and actin cytoskeleton compared to control cells (SNB19 Ctrl) (Figure 6). SNB19 TW cells formed far fewer and smaller cellular aggregates compared with control cells indicating a significant change in cell-cell interactions (Figure 6A). Cell substrate interaction, tested by cell plating on fibronectin (FN) showed over

100% greater adhesion of SNB19 TW cells than control cells compared with no significant change found when cells are plated on bovine serum albumin (Figure 6B). Migration of SNB19 TW cells through an uncoated filter membrane increased 40% compared with controls (Figure 6C). Consistent with increased migration, SNB19 TW cells showed reorganization of actin cytoskeleton with increased lamellipodia formation associated with activation of focal adhesion kinase (FAK) at leading edges (Figure 6D). Together, these results demonstrated a concordant relationship between the TWIST1-mediated program of gene expression and cellular phenotypes. Furthermore these data indicated that TWIST1 regulates the multiple individual cellular changes (alteration of cell-cell interaction, cell-substrate interactions and reorganization of actin cytoskeleton to facilitate migration) that together comprise the carefully orchestrated process of glioma invasion.

#### **TWIST1 activates expression of mesenchymal genes without cadherin switch**

The previous data demonstrated that TWIST1 over-expression promoted an invasive mesenchymal cellular

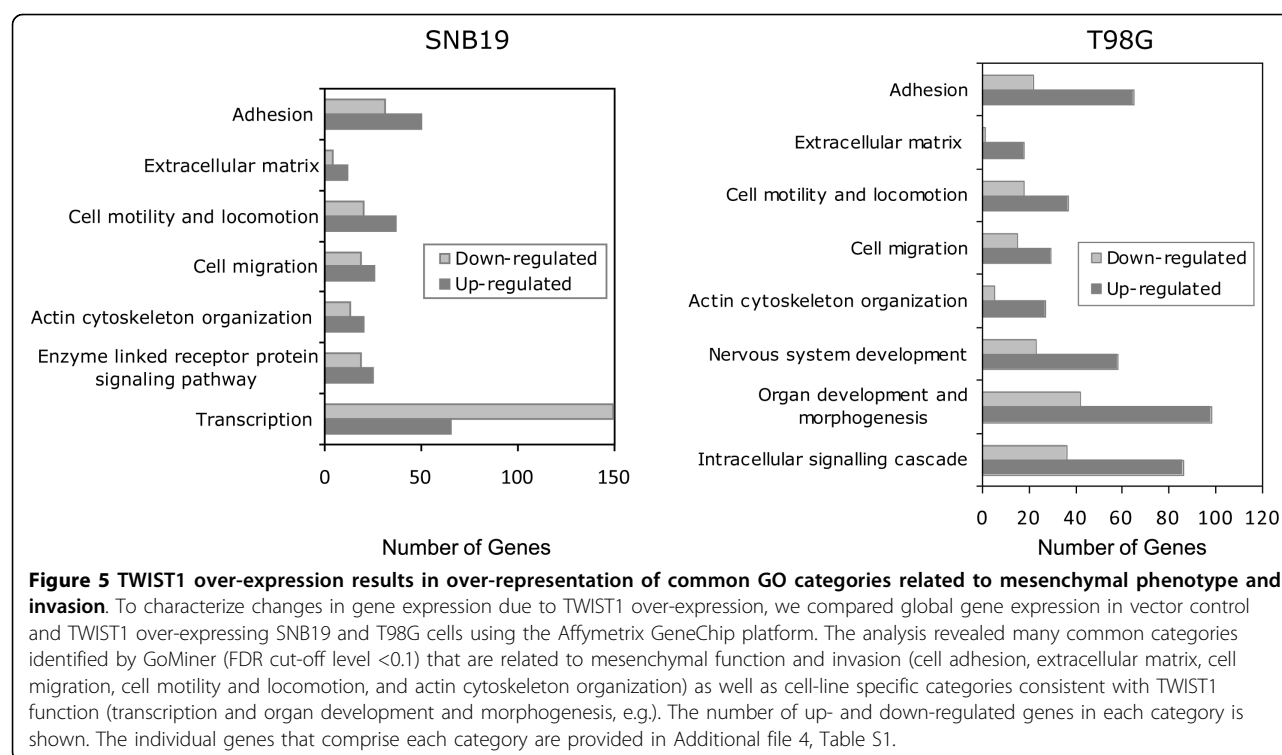


**Figure 4 Brightest point projection images (BPI) of tumor growth patterns.** The same tumors shown in Figure 3 were analyzed using BPI to visualize global differences in tumor growth patterns. Control T98G and T98G TWIST1 over-expressing tumor images are generated from total of 140 and 164 optical sections, respectively, collected by confocal microscope using ImageJ software. Control tumors possess a more cohesive pattern and localized growth pattern while T98G TW tumors demonstrate a markedly diffuse pattern of growth (type 3 invasive growth pattern). Arrows indicate regions of diffuse T98G TW tumor cell outgrowth.

phenotype in GBM cells. Given that TWIST1 promotes carcinoma invasion and metastasis through activation of EMT we sought to determine whether TWIST1 activated molecular features associated with carcinoma EMT in GBM cells. Within GO categories common to both SNB19 and T98G, approximately 1/3 of genes were upregulated in both cell lines while 2/3 were cell-type specific (Additional file 4, Table S1, S2). Many of these were genes known to promote mesenchymal changes in epithelial cancers (Table 1). Differential expression of a subset of genes selected from the arrays was validated by comparing expression in microarray with qRT-PCR (Figure 7). Genes associated with EMT were regulated by TWIST1 in SNB19 and/or T98G GBM cells including extracellular matrix proteins fibronectin 1 (FN1) [14], periostin (POSTN) [15] and SPARC

[16-18] protease MMP2 [19,20], transcription factor SNAI2 [21-23] transcriptional modifier ID1 [24], growth factor HGF [21,25] lysyl oxidase (LOX) [26] and cell adhesion protein cadherin 11 (CDH11) [27]. Other genes associated with mesenchymal phenotypes and glioma invasion, not yet formally linked to EMT, included laminin, alpha 4 (LAMA4) [28] and fibroblast activation protein alpha (FAP) [29].

Perhaps the most widely used marker for EMT in carcinomas is loss of E-cadherin and upregulation of N-cadherin, or the “cadherin switch”. To determine whether TWIST1 activated a “cadherin switch” in GBM cells, we quantified E-cadherin and N-cadherin mRNA expression in a panel of GBM cell lines each with vector control or TWIST1 over-expression. We found no consistent relationship between endogenous levels of E- or N-cadherin

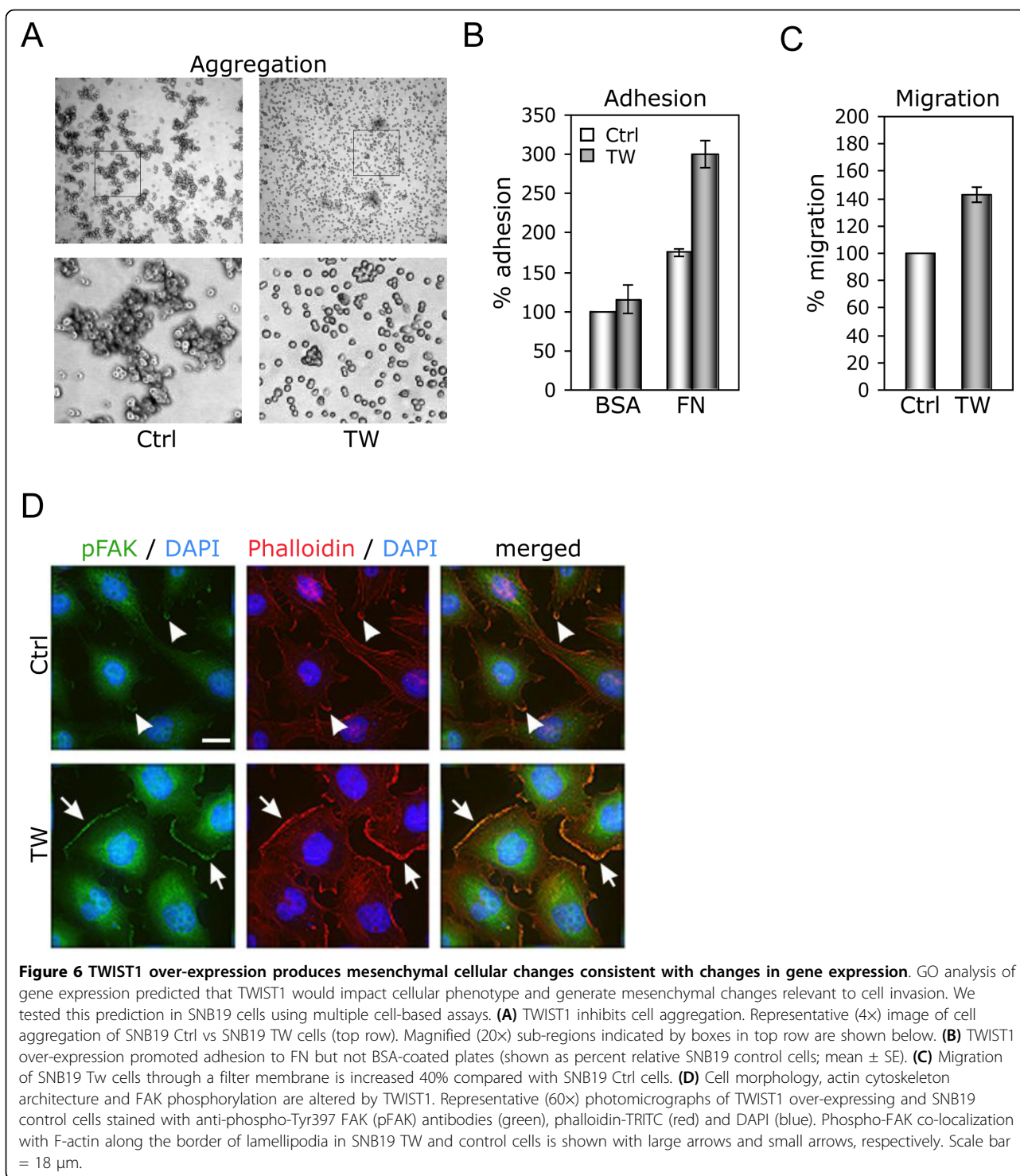


or changes with TWIST1 over-expression (Figure 8). Of importance, in no case did TWIST1 over-expression result in combined reduction of E-cadherin and upregulation of N-cadherin. These data indicated that the canonical “cadherin switch” central to TWIST1-mediated EMT in carcinomas does not occur in GBM cell lines over-expressing TWIST1 nor is it required to promote an invasive mesenchymal phenotype in human GBM cells. In only one of four cell lines (SNB19) in which TWIST1-mediated invasion was formally tested, did E-cadherin expression decrease with TWIST1 over-expression (Figure 8C). While TWIST1 over-expression resulted in decreased cell-cell adhesion in SNB19 in association with reduced E-cadherin expression, a similar decrease in cell-cell adhesion was noted for T98G TW cells versus vector controls (data not shown) despite the absence of detectable E-cadherin in control cells. Together these findings indicated that mechanisms independent of the E- to N-cadherin switch promote TWIST1-mediated GBM cell invasiveness. However, the commonality of genes regulated by TWIST1 in GBM cell lines and carcinoma metastasis suggested that the molecular program in GBM does partially overlap with that of TWIST1-mediated EMT in carcinomas.

#### TWIST1 expression highly correlates with *in vitro* TWIST1 target genes in human glioma samples

To determine whether expression of putative TWIST1 targets identified in microarrays are clinically relevant we

compared expression of SNAI2 (upregulated 6.6 fold in SNB19 TW cells) and FAP (upregulated 36 fold in SNB19 and 6.5 fold in T98G). These genes were selected for study because of their characterized roles in carcinoma EMT invasion and metastasis [30] and their reported upregulation in malignant gliomas [31,32]. In a set of 39 human glial neoplasms using quantitative RT-PCR we found a significant correlation between TWIST1 and SNAI2 ( $r = 0.72$ ;  $p = 0.001$ ) and TWIST1 and FAP alpha ( $r = 0.57$ ,  $p = 0.001$ ) message levels (Figure 9A, B). Compared to normal brain controls, the mean TWIST1, SNAI2 and FAP expression were all up-regulated in the most malignant grade IV gliomas compared with grade II and III tumors ( $p = 0.022$ ,  $p = 0.0014$ ,  $p = 0.005$ , respectively) (Figure 9C, D, E). Consistent with their roles in mesodermal development and mesenchymal differentiation, the highest expression levels for all genes were evident in grade IV gliosarcoma tumors, which have the highest degree of overt mesenchymal differentiation among all gliomas. These results demonstrated for the first time a close association between expression of TWIST1 and putative TWIST1 target genes also implicated in both carcinoma and glioma malignancy. To confirm that putative TWIST1 targets play a role in glioma cell invasiveness, we overexpressed SNAI2 in SNB19 cells and found that SNAI2 was sufficient to increase SNB19 cell invasion 80% (Additional file 5). Together our results support the relevance of our *in vitro* model of TWIST1 function to identify candidate mechanisms of TWIST1-mediated invasion.



#### Therapeutic significance of the inhibition of TWIST1 expression

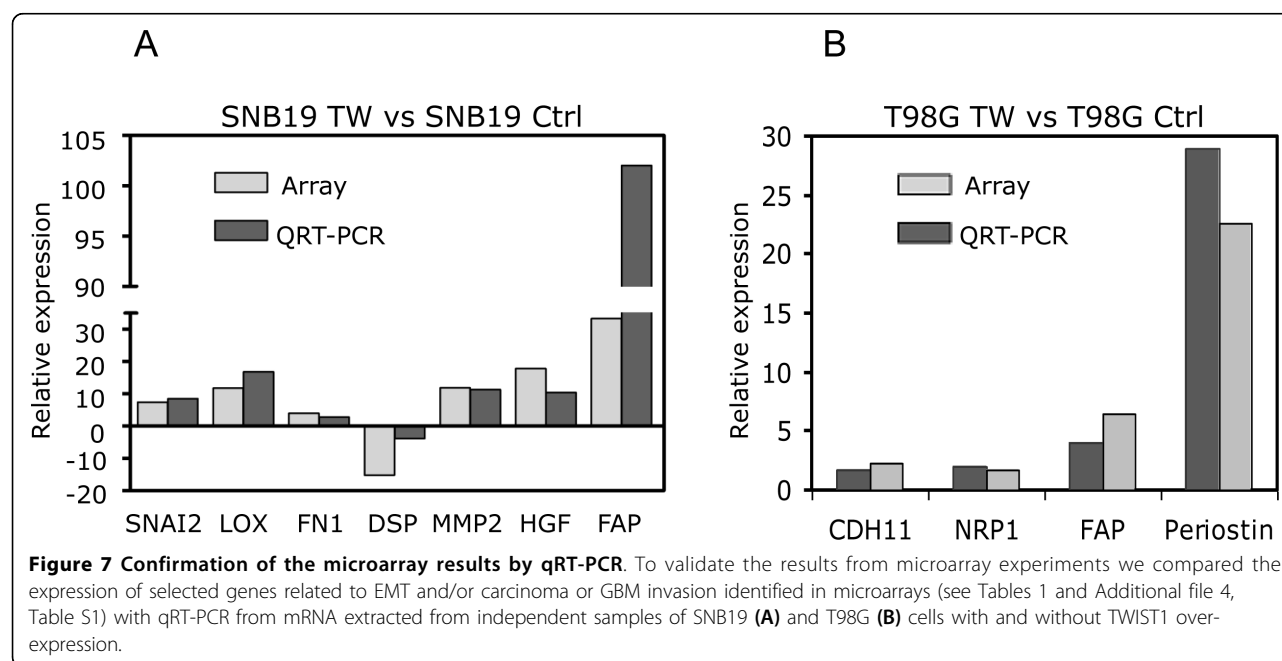
Our results demonstrated that increased levels of TWIST1 expression correlate with increased cell invasiveness. To determine whether inhibition of TWIST1 expression may have therapeutic relevance we

investigated the effects of TWIST1 knockdown on cell invasion and glioma stem cell proliferation as well as tumor sphere formation using sequence-specific shRNA lentiviral constructs. In SNB19 and T98G cell lines, decreasing levels of TWIST1 message and protein resulted in inhibition of invasion. For SNB19, a 40%



**Table 1 EMT and invasion genes regulated by TWIST1 in SNB19 and/or T98G GBM cell lines**

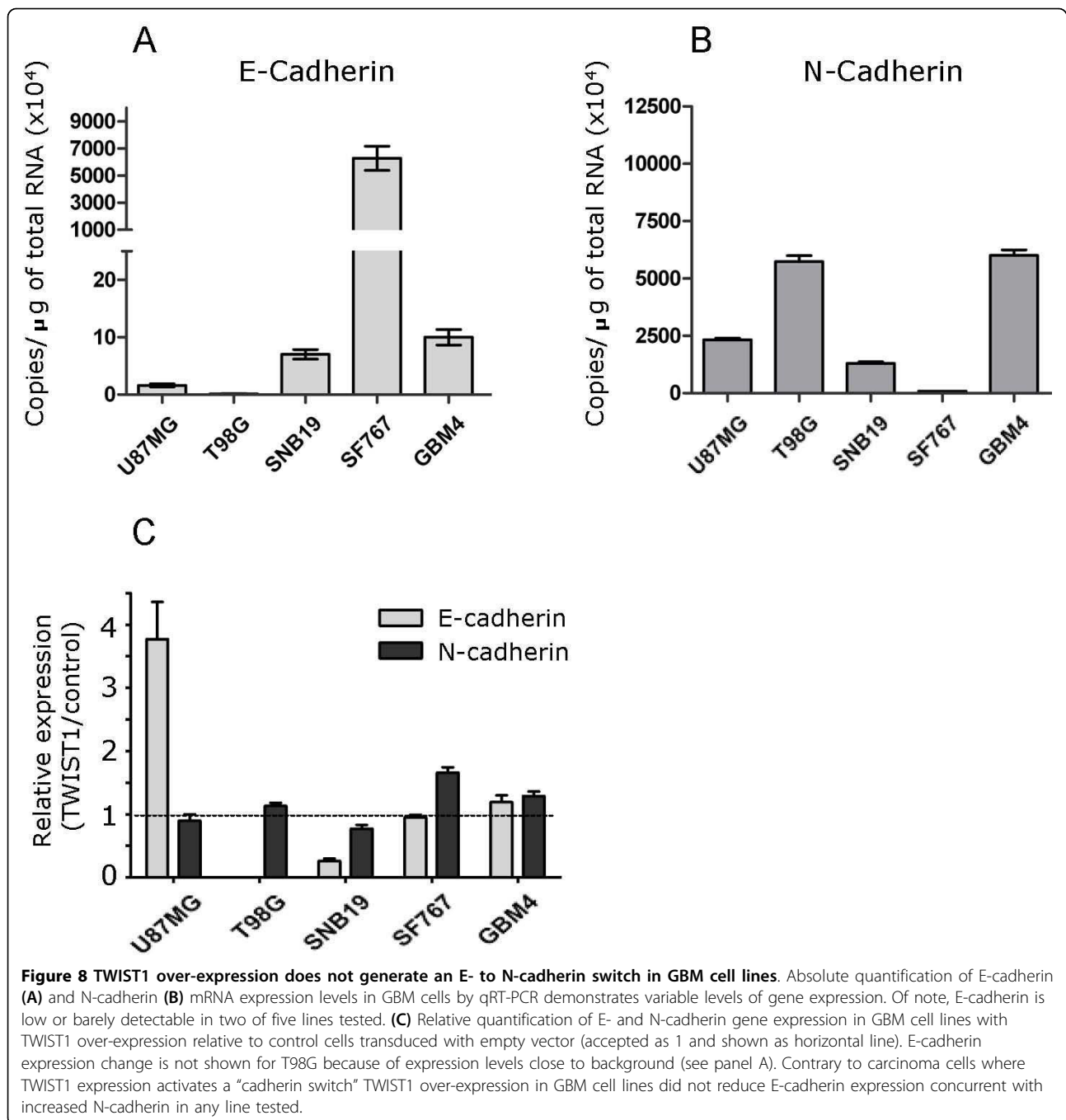
Gene symbol	Gene ID	Gene name	Fold differences	
			SNB19	T98G
SNAI2	213139_at	snail homolog 2	7.4	1
ID1	208937_s_at	inhibitor of DNA binding 1	2.0	2.1
ID2	213931_at	inhibitor of DNA binding 2	1.9	1.9
LOX	204298_s_at	lysyl oxidase	12	1
TIMP3	201149_s_at	TIMP metalloproteinase inhibitor 3	7.5	1
SPARC	212667_at	secreted protein, acidic, cysteine-rich	2.0	1
PDGFRB	202273_at	platelet-derived growth factor receptor	4.0	1
LAMA4	202202_s_at	laminin, alpha 4	7.7	4.8
FN1	216442_x_at	fibronectin 1	3.4	1
CDH11	236179_at	cadherin 11, type 2, OB-cadherin (osteoblast)	5.9	2.3
MMP2	201069_at	matrix metalloproteinase 2	12.7	1
FAP	209955_s_at	fibroblast activation protein, alpha	33.3	6.5
HGF	209960_at	hepatocyte growth factor	18	1
DSP	200606_at	Desmoplakin	-15.2	-3.8
IL8	211506_s_at	interleukin 8	7.5	12.1
POSTN	2105809_s_at	Periostin	1	22.8



suppression of TWIST1 mRNA (not shown) and concurrent reduction in protein levels reduced invasion 53% (Figure 10A) while in T98G cells an 80% reduction of TWIST1 mRNA levels (not shown) and concurrent reduction in protein levels resulted in 40% inhibition of invasion through matrigel (Figure 10B). Invasion was compared to control cells stably infected with shGFP lentiviral constructs. These controls did not affect TWIST1 expression or invasion compared with the parental cells not infected with shGFP lentivirus (data not

shown). A second, independent TWIST1-specific shRNA construct resulted in similar reductions of TWIST1 mRNA and invasion (data not shown). Finally, to demonstrate the specificity of the TWIST1 shRNAs we quantified their effects on the highly related gene Dermo1/TWIST2 and found no changes in its expression (data not shown). Using well-characterized glioma stem cells [12] we then tested the effect of TWIST1 inhibition on sphere-forming activity. Sphere number measures the frequency of cells capable of proliferating



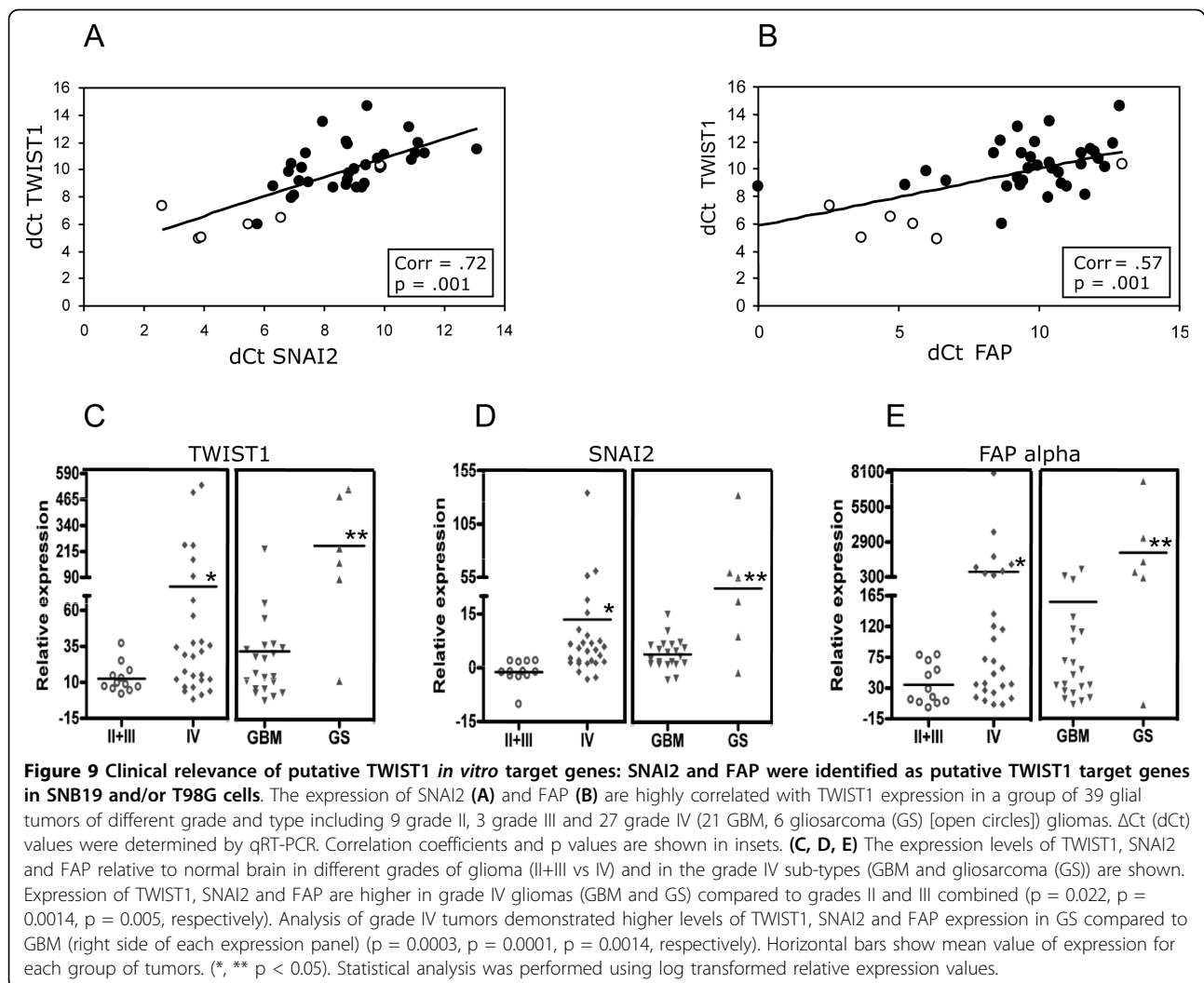


to form cell clusters while sphere size reflects the growth potential of each sphere. With 80% inhibition of TWIST1 expression in GBM6 stem cells, mean sphere size was reduced 48% ( $p < 0.0001$ ) (Figure 10 C-E). In the GBM8 tumor stem cell line, similar 80% inhibition of TWIST1 message (data not shown) resulted in a marked decrease in the number of wells that formed tumor spheres when plated at clonal dilution (30% vs. 67% in control GBM8 cells;  $p = 0.0033$ ) and the mean number of spheres in each well containing spheres (1.0

vs 2.3 for control GBM8 cells;  $p = 0.002$ ) (see Table 2). Together these results indicated the potential therapeutic relevance of TWIST1 inhibition for invasion and abrogation of glioma stem cell properties.

## Discussion

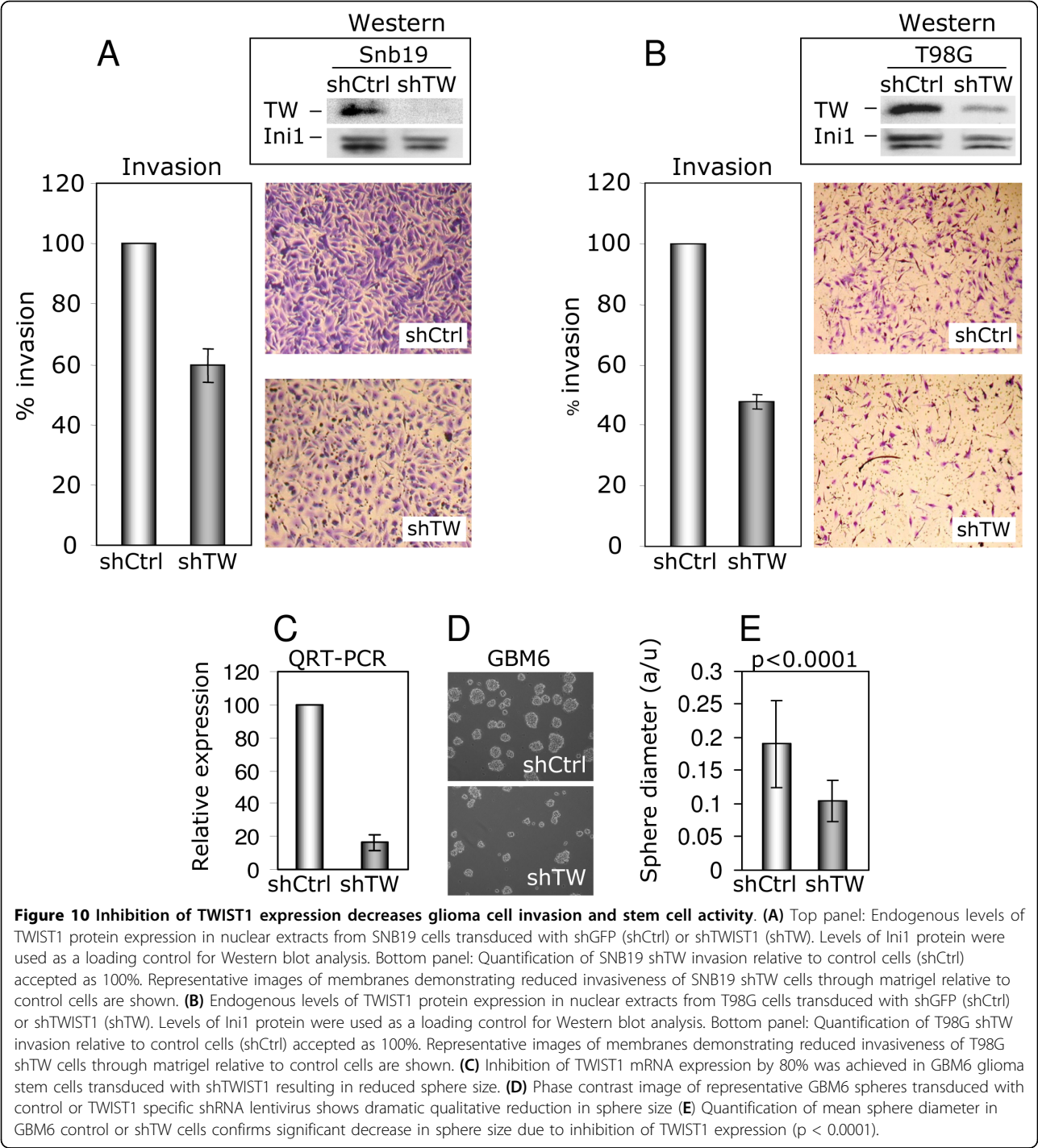
The fundamental role for mesenchymal change in promoting invasion, malignancy treatment response and even cancer stem cell function in human carcinoma and GBM invasion is increasingly recognized [8,9].



TWIST1 is a central regulator of mesenchymal change in carcinoma [5] but its relevance to invasion and mesenchymal change in GBM models has not been studied. Since tumor invasion is perhaps the major obstacle to improved outcome for patients with carcinomas and gliomas the elucidation of TWIST1 function in GBMs is potentially of great clinical importance. Following on our previous observation in the SF767 GBM cell line [11] this study validated the pro-invasive function of TWIST1 in multiple cell lines *in vitro* and *in vivo* and demonstrated that TWIST1 promoted clinically relevant mesenchymal molecular and cellular phenotypes that partially recapitulated those associated with carcinoma EMT. These findings identify TWIST1 as a regulator of mesenchymal change and invasion in GBM that can be leveraged for further investigation of the clinical potential of subverting mesenchymal change as a therapeutic strategy in treating GBM.

Collectively TWIST1 promoted invasion *in vitro* of all GBM cells tested to date (including a GBM stem cell line). We further established that TWIST1 enhanced invasion in the more relevant settings of brain slice culture and orthotopic xenotransplant models using SNB19 and T98G GBM cell lines. Of interest, the patterns of enhanced invasion generated by TWIST1 over-expression were cell-line specific with SNB19 TW cells invading as single cells or small aggregates from a central core while T98G TW cells diffusely invaded throughout the brain. These extreme patterns of invasion are similar to those in cases of gliomatosis cerebri [33]. These findings clearly demonstrate the generic pro-invasive function for TWIST1 in GBM and suggest that cell-intrinsic factors can modify TWIST1-mediated patterns of GBM invasion.

Consistent with this, TWIST1 over-expression generated cell-specific changes in gene expression with shared pro-invasive functional attributes. TWIST1-mediated



**Table 2 TWIST1 knockdown reduces sphere-forming activity of GBM8 stem cells**

Cell type	Percent wells with spheres	Number of spheres/well +/-SE
shCTRL	67	2.3+/-0.3
shTW	30*	1+/-0.1**

\*  $p = 0.0033$ ; \*\*  $p = 0.002$

changes in expression of specific genes in SNB19 and T98G were heterogeneous but overlapped at the functional level within five common categories related to the cellular requirements for glioma invasion and EMT including cell adhesion, extracellular matrix, cell motility and locomotion, cell migration and actin cytoskeleton organization. Importantly, TWIST1 over-expression

generated cell phenotypes highly consistent with the over-representation of genes within these functional categories that reflect critical individual cellular features required for carcinoma and GBM invasion [1]. We also determined that TWIST1 induced re-localization of activated FAK to sites of abundant lamellipodia formation, a significant finding given the association between FAK activation, cytoskeletal organization and its role in EMT and glioma malignancy (reviewed in [34-36]. Loss of apical-basal polarity (relative to a basement membrane) is an additional feature of EMT in carcinomas which was not tested here since assays of polarity for GBM cells *in vitro* are not well established. However, the recent description of polarized ciliated neural stem cells within the ventricular zone neuroepithelium [37-39] suggests that such studies could be attempted *in situ* or with novel co-culture systems or at earlier stages of glioma development. This approach could reveal polarity changes (analogous to carcinoma EMT) as fundamental steps in the process of gliomagenesis and acquisition of an invasive phenotype. Together gene expression analysis and cellular assays demonstrated that TWIST1 over-expression in glioma cells orchestrated the acquisition of a robust mesenchymal phenotype and cellular changes that closely mirror those of carcinoma cells undergoing mesenchymal transformation [40] and required for tumor invasion and metastasis [41].

TWIST1-mediated molecular changes also provided important insight into its role in mesenchymal change in GBM. Many genes related to carcinoma EMT were also up-regulated by TWIST1 in GBM indicating potential mechanistic overlap between the two processes. However, the lack of a TWIST1-mediated “cadherin switch” in GBM cells suggested that alternative mechanisms in nervous tissue and gliomas function to modulate cell adhesion and invasion. Alternatively, a cadherin switch could occur early in gliomagenesis or require specific anatomic or environmental interactions not present in our experimental system. The recent discovery that normal neural stem cells – putative GBM cells of origin – express E-cadherin supports this possibility [42,43]. Further studies are warranted to examine the impact of TWIST1 and other factors related to mesenchymal change in normal GBM cells of origin (neural stem and progenitor cells) or in cells at early stages of gliomagenesis to better define how alterations in E-cadherin or other cell-cell adhesion molecules impact the acquisition of an invasive malignant phenotype.

The clinical relevance of identified putative TWIST1 targets was established through correlation between TWIST1, SNAI2 and FAP expression levels in 39 human gliomas of different grades. These studies demonstrated that the current *in vitro* model of

TWIST1 pro-invasive function was capable of identifying clinically relevant pro-invasive targets and candidate downstream mechanisms of TWIST1-mediated glioma invasion. Our data also confirms prior reports that expression of SNAI2 [31] and FAP [44] is directly linked to malignant glioma grade and further showed that they are coordinately upregulated in gliosarcoma, the grade IV glioma with the most overt mesenchymal differentiation. As regulators of invasiveness, TWIST1 and SNAI2 are potential targets for therapeutic modulation, a proposition further supported by their known functions to promote cell survival and treatment resistance in other cancer types [45-50]. FAP is expressed in wounds and fibrotic tissues as well as carcinoma-associated fibroblasts in multiple cancer types and is thought to degrade tumor matrix and facilitate carcinoma invasion [51]. Further studies are needed to determine which cell type(s) express FAP and whether it serves a similar role of altering tumor stroma to promote invasion in GBM.

The significance of TWIST1 function to promote invasion through mesenchymal change in GBMs is underscored by recent reports of clinically relevant mesenchymal phenotypes in GBMs. Gene expression array studies identified a mesenchymal stem cell (MSC) phenotype in human GBMs [10] and distinct pro-neural, proliferative and mesenchymal gene expression signatures among malignant grade III and IV human gliomas [9]. The mesenchymal signature is associated with poor prognosis, increased angiogenesis and tumor recurrence [9]. Therefore, along with other transcription factors such as STAT3 and C/EBP which were recently identified as regulators of mesenchymal transformation in GBM cells [8] the correlation of TWIST1 with induction of mesenchymal changes, increased glioma grade and invasiveness implicate TWIST1 as an additional central regulator of this process in human GBM. Of note, STAT3 transcriptionally upregulates TWIST1 expression and promotes breast carcinoma cell migration [52] prompting speculation that STAT3-TWIST1 interactions in GBM may also contribute to invasion and mesenchymal change.

Inhibitors of TWIST1 are not available; therefore, to investigate the therapeutic relevance of inhibiting TWIST1 in GBM we knocked down TWIST1 expression using shRNA and assayed its effects on cell invasion and glioma stem cell properties. Specific inhibition of TWIST expression resulted in marked reductions in glioma cell invasion *in vitro*. These findings are consistent with the pro-invasive function of TWIST1 in GBM and support the therapeutic potential of inhibiting TWIST1 or TWIST1-mediated signaling to inhibit GBM invasion. Glioma stem cells are recognized as tumor-initiating cells that determine tumor malignancy and growth. Through activation of EMT, TWIST1

promotes the formation and maintenance of breast cancer stem cells [53] and TWIST1 over-expression is implicated in mesenchymal stem cell activity [54]. Given these observations we propose that targeting TWIST1 may have additional therapeutic relevance in gliomas by abrogating glioma stem cell functions. Our data showed that inhibition of TWIST1 expression resulted in a dramatic reduction in GBM stem cell sphere formation and growth. These results suggest that a unique therapeutic potential of inhibiting TWIST1 may result from simultaneous targeting of glioma cell invasiveness and stem cell function – hallmark GBM properties that both contribute to tumor growth, progression and treatment resistance. To address this potential, ongoing and future studies will address the effects of TWIST1 inhibition in GBM cells on tumor growth, invasion and response to therapy *in vivo*.

## Conclusions

Together these studies demonstrated that TWIST1 enhances GBM invasion in concert with mesenchymal changes. However, these changes do not involve the canonical cadherin switch of carcinoma EMT. The present findings demonstrated the potential usefulness of applying carcinoma EMT as a framework from which to enhance our understanding of GBM invasion and further suggest that a neural form of mesenchymal change, analogous to carcinoma EMT, may contribute broadly to glioma malignancy. Based on these findings we propose targeting TWIST1-mediated mesenchymal change as a therapeutic strategy with potential to inhibit GBM invasion and tumor growth, and enhance treatment responses.

## Methods

### Cell lines and tissue

Glioblastoma cell lines T98G, SNB19, SF767, U87MG were maintained in DMEM/F12 with 10% FBS (Hyclone). Human primary GBM cancer-initiated cells (GBM4, GBM6) were cultured as described [12] in the presence of EGF and bFGF. Human glioma tumor samples were acquired according to a protocol approved by the Institutional Review Board of the Human Subjects Division of the University of Washington. Samples were immediately snap frozen in liquid nitrogen and stored at -80°C before processing. Type and grade of tumors were confirmed by histopathological examination.

### Expression and shRNA constructs, and cell transduction

A retroviral human TWIST over-expression construct and methods for infection of SNB19 and T98G cell lines were described previously [11]. Myc-tagged SNAI2 expression construct (SNAI2myc) was generated by PCR followed by subcloning in LXS expression vector. Exogenous protein expression was confirmed by Western

blot analyses with corresponding antibody. Lentiviral shRNA construct for inhibition of TWIST1 expression [5] and control shRNA were purchased from Addgene. Lentivirus was generated using a standard method in HEK293T cells. A pool of infected cells was selected with Puromycin (1 µg/mL).

### Cell aggregation assay

Single-cell suspensions ( $10^5$  cells/mL in DMEM-F12 without FBS) were plated into each well of a 6-well plate coated with 0.6% agarose/DMEM-F12. The plate was incubated at 37°C on a rocking platform for 16 hrs. Cells were fixed in 5% formalin to preserve cell-cell interactions and photographed.

### Cell adhesion

SNB19 LXS and TWIST cells ( $5 \times 10^4$ /1 mL) were allowed to adhere to 24-well plates coated with BSA or fibronectin (5 µg/mL) for 1 hour. Cells were then washed, fixed, stained and counted. Differences in cell adhesion are shown as percent of SNB19 control cells attached to BSA-coated wells. Three wells from 3 separate experiments were analyzed and the significance of differences was determined by Student t-test. Data shown are mean  $\pm$  SE.

### GBM stem cell sphere assays

GBM6 and GBM8 stem cells with TWIST1 knockdown and control cells (scrambled shRNA lentiviral vector) were dissociated and viable cells were counted using ViCell. Viable GBM6 cells plated at 3200 cells per well in 6-well plates were used to establish the effect of TWIST1 knockdown on sphere size. After 5 days spheres were photographed and sphere sizes were measured using Adobe Photoshop. To determine the effects of TWIST1 on sphere-forming activity GBM8 cells were plated at clonal dilution (20 viable cells per well in 96-well plate). After 7 days, wells with spheres were counted and presented as a percent of wells with spheres. Average numbers of spheres per well with spheres were also calculated. Fisher exact test and t-test were used for statistical analysis as appropriate.

### Immunocytochemistry and F-actin staining

Cells were grown in 8-well chambers and fixed in 4% PFA for 10 min. Following treatment with 0.1% Triton X-100/TBS for 5 min, cells were blocked in 1%BSA/TBS, washed and incubated with FAK or phospho-FAK antibody (Upstate) according to manufacturer protocol. Appropriate secondary antibody conjugated with FITC (Pierce) was used for antigen detection. F-actin was stained with TRITC conjugated phalloidin (1 µg/mL) followed by DAPI (1 µg/mL) staining. Antigens were visualized using confocal microscopy (Delta-Vision).

### Western blot analysis

Cell harvesting, cell lysis and Western blot procedures were performed as described previously [11]. Total cell lysates were used for detection of TWIST1 over-expression. Nuclear extracts were used to increase assay sensitivity in detecting endogenous TWIST1 expression. For protein loading control, anti- $\beta$ -actin antibody (Sigma) for total proteins or Ini1 (H-300) antibody (Santa Cruz) for nuclear proteins were used. Immunoblot for MycTag Antibody (Upstate) was performed according to manufacturer's recommendation.

### Invasion and migration assays

The invasion and migration assays were performed using 24-well Matrigel invasion chambers or uncoated Control inserts (BD Biosciences) as previously described [11]. Briefly, cells were resuspended in a serum-free DMEM and loaded into inserts ( $5 \times 10^4$  cells/500  $\mu$ L). DMEM/F-12 with 10% FBS (SNB19) and without FBS (T98G) was added to the lower chamber (750  $\mu$ L). Following incubation at 37°C, cells that invaded or migrated to the underside of the membrane were fixed, stained, digitally imaged and counted. Differences in cell invasion were expressed as a percent of invading/migrating relative to control cells. Data shown are mean  $\pm$  SE.

### Invasion in organotypic brain slices

Coronal brain slices (400  $\mu$ m) from 21-day-old mice were cultured as previously described in the media supplemented with 10%FBS [55]. SNB19 or T98G cells with LXS or TWIST1 over-expression (labeled with GFP expressing lentivirus) were placed at the corpus callosum. After 10 days *in vitro*, the co-cultures were fixed and analyzed by confocal microscopy. Slices were imaged using a FluoView FV1000 confocal microscope (Olympus). Collected Z-stacks were processed for visualization and cell counting using NIH Image software. The morphometric software Metamorph (Molecular Devices Corporation) was used to measure cell migration. The distances from the border of the cells' aggregate to each of the 20 furthest cells were measured for each of the 9 tissue slices (total  $n = 178$  measurements) and compared using a repeated measures ANOVA model. The modeling was done with the 'proc mixed' procedure available in SAS.

### Intracranial injection and whole brain imaging

Animal experiments were performed according to procedures approved by the University of Washington IACUC. Glioma cells harboring empty vector and cells with TWIST1 over-expression were labeled with GFP-expressing lentivirus (pLL3.7) prior to implantation. T98G cells were injected in 9- to 10-week-old SCID-NOD mice. SNB19 cells were injected in 7- to 8-week-old nude mice.

Following animal sedation  $3 \times 10^5$  labeled cells were injected intracranially into the right caudate nucleus using a stereotactic apparatus and a Hamilton syringe. To determine the effect of TWIST1 on tumor growth and invasion, animals (6 mice with SNB19 TW and 4 mice with SNB19 Ctrl) were sacrificed 17 days after injection when the animals first showed signs of morbidity. Animals injected with T98G TW or control cells (3 mice per group) were sacrificed 90 days after injection. Animals were perfused with 4% paraformaldehyde (PFA). The entire brain was dissected from the calvarial vault and fixed for an additional 24 hours in 4% PFA at room temperature with light agitation. Brains were washed with PBS and transferred to 50% glycerol in PBS for 24 hours, 75% glycerol for 24 hours, then 90% glycerol at 4°C until imaged. The whole brain was sliced in the axial plane to obtain 3 slices, each approximately 2 mm thick. Each slice then was imaged using the FV1000 laser scanning confocal microscope in the axial plane to detect GFP-expressing cells and perform automated image splicing to reconstruct the entire tumor in a single axial slice.

### Image analysis for invasion

A semi-quantitative scale was used for initial characterization of tumor growth patterns as follows: Type 1 = solid compact core with mainly localized expansile growth; Type 2 = easily detectable non-contiguous individual cells or cell clusters invading into brain parenchyma adjacent to the solid tumor core; Type 3 = poorly defined or absent core with diffusely invasive contiguous or non-contiguous growth. Type 3 represents the most invasive growth pattern. For circumstances where the scale did not provide clear indications for the degree of invasiveness, differences in tumor growth pattern were quantified as follows: Reconstructed wide-field brain images with tumors were analyzed using Huygens software (Scientific Volume Imaging, Hilversum, The Netherlands). Individual signal intensities from each individual optical section (Z-stack images) collected by confocal microscopy were integrated into a brightest point projection image (BPI) to provide a 2-dimensional summary of total tumor cell density and spatial distributions (Image J). Information collected includes tumor core volume, invasive cell volume and a number of invasive aggregates. Numerical values were compared using t-test.

### Microarray processing methods for the Affymetrix microarray platform

Gene expression experiments were performed using the GeneChip platform by Affymetrix (Santa Clara, CA) and the manufacturer's protocol. Statistical analysis and data normalization for the Affymetrix arrays were carried out



with Bioconductor software [56], and GeneTraffic® (Iobion Informatics LLC, La Jolla, CA). Modified t-test was applied for two-group comparison. Bioconductor was used to calculate p-values using a modified t-test in conjunction with an empirical Bayes method to moderate the standard errors of the estimated log-fold changes. P-values were adjusted for multiplicity with the program q-value. Genes with absolute change greater than or equal to 1.5 fold and  $p < 0.05$  were considered differentially regulated by TWIST1. To demonstrate the correlation of changes in expression (up or down-regulation) between genes differentially regulated in both cell lines, we applied a Pearson's correlation analysis using Bioconductor software (see above). Gene Ontology (GO) categories were analyzed using GoMiner [57] to detect gene category over-representation with cut off false discovery rate (FDR) 0.1.

#### Quantitative RT-PCR (qRT-PCR)

Total RNA from cells, brain or tumor samples was extracted using Qiagen RNeasy mini kit. RNA (1 µg) was reverse-transcribed with Clontech kit. SYBR Green PCR Master mix (ABI) was used for template amplification. Thermocycling for all targets was carried out in 30 µL reaction for 40 cycles in triplicate. Each cycle consisted of: 94°C for 15 seconds, 58°C for 30 seconds and 72°C for 30 seconds. For all samples, reactions were run in triplicate. PCR reactions where reverse transcriptase was omitted were used as negative controls. SYBR Green incorporation was monitored in real time with an ABI PRISM 7000 sequence detection system (Applied Biosystems) and threshold exponential amplification cycle ( $C_T$ ) was calculated by SDS system software. Differences in the  $C_T$  values ( $\Delta C_T$ ) between the target transcript and GAPDH endogenous control determined the relative gene expression level and the  $\Delta\Delta C_T$  method was used to calculate fold differences in expression. Relative expression in each tumor sample is normalized by expression of corresponding target in a pool of normal brain samples ( $n = 4$ ). Correlations between TWIST1 and SNAI2 or FAP expression were calculated using a regression coefficient and  $\Delta C_T$  values for each tumor sample. Statistical analysis of relative expression levels in human tumors was performed using unpaired t-test. Relative expression values were log transformed before testing to ensure normal distribution. Absolute quantification was used to compare levels of E- and N-cadherin mRNA expression. Standard curve was built using plasmid harboring corresponding cDNA targets diluted from 0 to  $10^7$  copies per reaction. Final results are shown as a number of copies per µg of total RNA. The specificity of amplifications was confirmed by amplicon melting profile.

## Additional material

### Additional file 1: Alteration of TWIST1 expression correlates with cell invasiveness *in vitro* in SNB19 and T98G cells. (A)

Top panel: Detection of TWIST1 protein from whole cell lysates of a pool of SNB19 cells transduced with retroviral expression construct (TW) compared to SNB19 cells transduced with empty vector (Ctrl).  $\beta$ -Actin is shown as loading control. Bottom panel: Quantification of SNB19 Tw cell invasion relative to Ctrl cells accepted as 100%. Representative images of membranes demonstrating increased invasiveness of Snb19 Tw cells relative to control cells are shown. (B) Top panel: Detection of exogenous TWIST1 over-expression in cell lysates from T98G cells by Western blot. Bottom panel: Quantification of T98G Tw cell invasion relative to Ctrl cells accepted as 100%. Representative images of membranes demonstrating increased invasiveness of T98G Tw cells relative to control cells are shown.

### Additional file 2: Over-expression of TWIST1 expression in GBM stem cells correlates with cell invasiveness (A)

Quantification of exogenous TWIST1 over-expression using qRT-PCR in GBM4 primary GBM stem cells cultured as neurospheres and transduced with TWIST1 retroviral expression vector. (B) Quantification of GBM4 cell invasiveness in matrigel assay. Representative images of membranes demonstrating increased invasiveness of GBM4 Tw cells relative to control are shown. Differences in cell invasion are shown as percent of control cells transduced with empty vector (mean  $\pm$  SE).

### Additional file 3: Pearson's correlation of genes differentially regulated by TWIST1 in T98G and SNB19 cells with TWIST1 over-expression relative to corresponding controls. A total of 189 genes (1.5 fold, $p < 0.05$ ) were differentially co-regulated by TWIST1 in both T98G and SNB19 cells relative to corresponding controls.

**Additional file 4: Supplementary Tables. Table S1.** The list of common and cell-specific differentially expressed genes within each GO category over-represented in both SNB19 and T98G cells. **Table S2.** The total number of genes within common categories shown in Table S1.

### Additional file 5: Putative TWIST1 target SNAI2 is sufficient to induce glioma invasiveness *in vitro*. (A)

Exogenous over-expression of Myc-tagged SNAI2 in SNB19 cells. (B) Quantification of invasion of SNB19 cells with SNAI2 over-expression. Representative images of invasive cells on the membrane are shown.

## Abbreviations

EGF: epidermal growth factor; bFGF: basic fibroblast growth factor; FAP: fibroblast activating protein alpha; FAK: focal adhesion kinase; SNAI2: snail homolog 2; MMP2: Matrix metalloproteinase-2; HGF: hepatocyte growth factor; LAMA4: laminin alpha 4; LOX: lysyl oxidase; ADAM12: A disintegrin and metalloproteinase domain 12; DSP: desmoplakin; FBS: fetal bovine serum.

## Acknowledgements and Financial Support

We acknowledge Rosemary Kimmel for her expert editorial assistance, J. Barber for the help with statistical analysis, Drs Jing Zhang, Eduardo Mendez and Daniel Silbergeld for critical review of the manuscript, Theo Bammler and Frederico Farin of the UW CHDD Genomics Core for assistance with microarray studies, Glen MacDonald of the CHDD Cellular Morphology Core for assistance with development of whole brain tumor imaging techniques, Drs Daniel L. Silbergeld, Alexander M. Spence, Jason Rockhill, and Maciej Mrugala whose clinical efforts made this work possible. This research was funded in part through an NIH/NINDS T32-NS-0007144 Clinical Neuroscience Training Grant (RO, LK, JM) and a University of Washington Institutional Bridge Funding Grant (RCR, AKM). Research in ISG group is supported partially by FEDER and by MICINN (SAF2009-08803), Junta de Castilla y León (CSI13A08 and proyecto Biomedicina 2009-2010), MEC OncoBIO Consolider-Ingenio 2010 (Ref. CSD2007-0017), Sandra Ibarra Foundation, NIH grant (R01 CA109335-04A1) and by Group of Excellence Grant (GR15) from Junta de Castilla y León. There are no conflicts of interest.

## Author details

<sup>1</sup>Department of Neurological Surgery, University of Washington School of Medicine, Seattle WA, 98195, USA. <sup>2</sup>Environmental Health Sciences, University of Washington School of Medicine, Seattle WA, 98195, USA. <sup>3</sup>Institute for Stem Cell and Regenerative Medicine, University of Washington School of Medicine, Seattle WA, 98195 USA. <sup>4</sup>Division of Molecular Medicine, Beckman Research Institute of the City of Hope, Duarte CA, 91010, USA. <sup>5</sup>Molecular Neurosurgery Laboratory, Massachusetts General Hospital, Harvard Medical School, Boston MA, 02114 USA. <sup>6</sup>Experimental Therapeutics and Translational Oncology Program, Instituto de Biología Molecular y Celular del Cáncer (IBMCC), CSIC/Universidad de Salamanca, Campus Unamuno, 37007-Salamanca, Spain. <sup>7</sup>Petrov Research Institute of Oncology, St.-Petersburg, 197758, Russia.

## Authors' contributions

SAM generated expression constructs and performed *in vitro* cellular assays, qRT-PCR and microarrays. AMM carried out *in vivo* studies, confocal imaging, microarray interpretation and statistical data analysis. AP performed *ex vivo* invasion assays. RB performed microarray analysis and bioinformatics support. RGO and LK participated in tumor collection and assisted with cell culture experiments. JPM participated in generation of recombinant DNAs and analysis of gene over-expression and knockdown. CAG generated anti-TWIST1 antibody. HW generated primary GBM cells. IG-H and IS-G generated reagents for Snai2 analysis. JRS and PJH assisted in experiment design, data interpretation and manuscript writing. RCR conceived of the study, and participated in its design, coordination and manuscript writing. All authors read and approved the final manuscript.

## Competing interests

The authors declare that they have no competing interests.

Received: 23 February 2010 Accepted: 20 July 2010

Published: 20 July 2010

## References

- Kalluri R, Weinberg RA: **The basics of epithelial-mesenchymal transition.** *J Clin Invest* 2009, **119**:1420-1428.
- Yu W, Kamara H, Svoboda KK: **The role of twist during palate development.** *Dev Dyn* 2008, **237**:2716-2725.
- Venkov CD, Link AJ, Jennings JL, Plieth D, Inoue T, Nagai K, Xu C, Dimitrova YN, Rauscher FJ, Neilson EG: **A proximal activator of transcription in epithelial-mesenchymal transition.** *J Clin Invest* 2007, **117**:482-491.
- Bridges RS, Kass D, Loh K, Glackin C, Borczuk AC, Greenberg S: **Gene expression profiling of pulmonary fibrosis identifies Twist1 as an antiapoptotic molecular "rectifier" of growth factor signaling.** *Am J Pathol* 2009, **175**:2351-2361.
- Yang J, Mani SA, Donaher JL, Ramaswamy S, Itzykson RA, Come C, Savagner P, Gitelman I, Richardson A, Weinberg RA: **Twist, a master regulator of morphogenesis, plays an essential role in tumor metastasis.** *Cell* 2004, **117**:927-939.
- Yang MH, Wu MZ, Chiou SH, Chen PM, Chang SY, Liu CJ, Teng SC, Wu KJ: **Direct regulation of TWIST by HIF-1alpha promotes metastasis.** *Nat Cell Biol* 2008, **10**:295-305.
- Yang Z, Zhang X, Gang H, Li X, Li Z, Wang T, Han J, Luo T, Wen F, Wu X: **Up-regulation of gastric cancer cell invasion by Twist is accompanied by N-cadherin and fibronectin expression.** *Biochem Biophys Res Commun* 2007, **358**:925-930.
- Carro MS, Lim WK, Alvarez MJ, Bollo RJ, Zhao X, Snyder EY, Sulman EP, Anne SL, Doetsch F, Colman H, Lasorella A, Aldape K, Califano A, Iavarone A: **The transcriptional network for mesenchymal transformation of brain tumours.** *Nature* 2010, **463**:318-325.
- Phillips HS, Kharbanda S, Chen R, Forrest WF, Soriano RH, Wu TD, Misra A, Nigro JM, Colman H, Soroceanu L, Williams PM, Modrusan Z, Feuerstein BG, Aldape K: **Molecular subclasses of high-grade glioma predict prognosis, delineate a pattern of disease progression, and resemble stages in neurogenesis.** *Cancer Cell* 2006, **9**:157-173.
- Tso CL, Shintaku P, Chen J, Liu Q, Liu J, Chen Z, Yoshimoto K, Mischel PS, Cloughesy TF, Liaw LM, Nelson SF: **Primary glioblastomas express mesenchymal stem-like properties.** *Mol Cancer Res* 2006, **4**:607-619.
- Elias MC, Tozer KR, Silber JR, Mikheeva S, Deng M, Morrison RS, Manning TC, Silbergeld DL, Glackin CA, Reh TA, Rostomily RC: **TWIST is expressed in human gliomas and promotes invasion.** *Neoplasia* 2005, **7**:824-837.
- Wakimoto H, Kesari S, Farrell CJ, Curry WT Jr, Zaupa C, Aghi M, Kuroda T, Stemmer-Rachamimov A, Shah K, Liu TC, Jeyaretna DS, Debasitis J, Pruszak J, Martuza RL, Rabkin SD: **Human glioblastoma-derived cancer stem cells: establishment of invasive glioma models and treatment with oncolytic herpes simplex virus vectors.** *Cancer Res* 2009, **69**:3472-3481.
- Kalluri R: **EMT: when epithelial cells decide to become mesenchymal-like cells.** *J Clin Invest* 2009, **119**:1417-1419.
- Zeisberg M, Neilson EG: **Biomarkers for epithelial-mesenchymal transitions.** *J Clin Invest* 2009, **119**:1429-1437.
- Ruan K, Bao S, Ouyang G: **The multifaceted role of periostin in tumorigenesis.** *Cell Mol Life Sci* 2009, **66**:2219-2230.
- Mathias RA, Chen YS, Wang B, Ji H, Kapp EA, Moritz RL, Zhu HJ, Simpson RJ: **Extracellular Remodelling During Oncogenic Ras-Induced Epithelial-Mesenchymal Transition Facilitates MDCK Cell Migration.** *J Proteome Res* 2010, **9**:1007-1019.
- Sario D, Rodriguez-Pinilla SM, Hardisson D, Cano A, Moreno-Bueno G, Palacios J: **Epithelial-mesenchymal transition in breast cancer relates to the basal-like phenotype.** *Cancer Res* 2008, **68**:989-997.
- Shi Q, Bao S, Song L, Wu Q, Bigner DD, Hjelmeland AB, Rich JN: **Targeting SPARC expression decreases glioma cellular survival and invasion associated with reduced activities of FAK and ILK kinases.** *Oncogene* 2007, **26**:4084-4094.
- Rupp PA, Visconti RP, Czirok A, Cheresh DA, Little CD: **Matrix metalloproteinase 2-integrin alpha(v)beta3 binding is required for mesenchymal cell invasive activity but not epithelial locomotion: a computational time-lapse study.** *Mol Biol Cell* 2008, **19**:5529-5540.
- Fillmore HL, VanMeter TE, Broadus WC: **Membrane-type matrix metalloproteinases (MT-MMPs): expression and function during glioma invasion.** *J Neurooncol* 2001, **53**:187-202.
- Savagner P, Yamada KM, Thiery JP: **The zinc-finger protein slug causes desmosome dissociation, an initial and necessary step for growth factor-induced epithelial-mesenchymal transition.** *J Cell Biol* 1997, **137**:1403-1419.
- Alves CC, Carneiro F, Hoefler H, Becker KF: **Role of the epithelial-mesenchymal transition regulator Slug in primary human cancers.** *Front Biosci* 2009, **14**:3035-3050.
- Cobaleda C, Perez-Caro M, Vicente-Duenas C, Sanchez-Garcia I: **Function of the zinc-finger transcription factor SNAI2 in cancer and development.** *Annu Rev Genet* 2007, **41**:41-61.
- Li Y, Yang J, Luo JH, Dedhar S, Liu Y: **Tubular epithelial cell dedifferentiation is driven by the helix-loop-helix transcriptional inhibitor Id1.** *J Am Soc Nephrol* 2007, **18**:449-460.
- Vergara D, Merlot B, Lucot JP, Collinet P, Vinatier D, Fournier I, Salzet M: **Epithelial-mesenchymal transition in ovarian cancer.** *Cancer Lett* 2009, **291**:59-66.
- Schietke RE, Warnecke C, Wacker I, Schodel J, Mole DR, Campean V, Amann K, Goppelt-Strube M, Behrens J, Eckardt KU, Wiesener MS: **The lysyl oxidases LOX and LOXL2 are necessary and sufficient to repress E-cadherin in hypoxia - insights into cellular transformation processes mediated by HIF-1.** *J Biol Chem* 2010, **285**:6658-6669.
- Simonneau L, Kitagawa M, Suzuki S, Thiery JP: **Cadherin 11 expression marks the mesenchymal phenotype: towards new functions for cadherins?** *Cell Adhes Commun* 1995, **3**:115-130.
- Nagato S, Nakagawa K, Harada H, Kohno S, Fujiwara H, Sekiguchi K, Ohue S, Iwata S, Ohnishi T: **Downregulation of laminin alpha4 chain expression inhibits glioma invasion in vitro and in vivo.** *Int J Cancer* 2005, **117**:41-50.
- Huang Y, Wang S, Kelly T: **Seprase promotes rapid tumor growth and increased microvessel density in a mouse model of human breast cancer.** *Cancer Res* 2004, **64**:2712-2716.
- Henry LR, Lee HO, Lee JS, Klein-Szanto A, Watts P, Ross EA, Chen WT, Cheng JD: **Clinical implications of fibroblast activation protein in patients with colon cancer.** *Clin Cancer Res* 2007, **13**:1736-1741.
- Scrideli CA, Carloti CG Jr, Okamoto OK, Andrade VS, Cortez MA, Motta FJ, Lucio-Eterovic AK, Neder L, Rosemberg S, Oba-Shinjo SM, Marie SK, Tone LG: **Gene expression profile analysis of primary glioblastomas and non-neoplastic brain tissue: identification of potential target genes by oligonucleotide microarray and real-time quantitative PCR.** *J Neurooncol* 2008, **88**:281-291.

32. Yamoutpour F, Bodempudi V, Park SE, Pan W, Mauzy MJ, Kratzke RA, Dudek A, Potter DA, Woo RA, O'Rourke DM, Tindall DJ, Farassati F: **Gene silencing for epidermal growth factor receptor variant III induces cell-specific cytotoxicity.** *Mol Cancer Ther* 2008, **7**:3586-3597.
33. Park S, Suh YL, Nam DH, Kim ST: **Gliomatosis cerebri: clinicopathologic study of 33 cases and comparison of mass forming and diffuse types.** *Clin Neuropathol* 2009, **28**:73-82.
34. Gunther W, Skaftnesmo KO, Arnold H, Terzis AJ: **Molecular approaches to brain tumour invasion.** *Acta Neurochir (Wien)* 2003, **145**:1029-1036.
35. Natarajan M, Hecker TP, Gladson CL: **FAK signaling in anaplastic astrocytoma and glioblastoma tumors.** *Cancer J* 2003, **9**:126-133.
36. Gutenberg A, Bruck W, Buchfelder M, Ludwig HC: **Expression of tyrosine kinases FAK and Pyk2 in 331 human astrocytomas.** *Acta Neuropathol (Berl)* 2004, **108**:224-230.
37. Chojnacki AK, Mak GK, Weiss S: **Identity crisis for adult periventricular neural stem cells: subventricular zone astrocytes, ependymal cells or both?** *Nat Rev Neurosci* 2009, **10**:153-163.
38. Mirzadeh Z, Merkle FT, Soriano-Navarro M, Garcia-Verdugo JM, Alvarez-Buylla A: **Neural stem cells confer unique pinwheel architecture to the ventricular surface in neurogenic regions of the adult brain.** *Cell Stem Cell* 2008, **3**:265-278.
39. Shen Q, Wang Y, Kokovay E, Lin G, Chuang SM, Goderie SK, Roysam B, Temple S: **Adult SVZ stem cells lie in a vascular niche: a quantitative analysis of niche cell-cell interactions.** *Cell Stem Cell* 2008, **3**:289-300.
40. Thiery JP, Sleeman JP: **Complex networks orchestrate epithelial-mesenchymal transitions.** *Nat Rev Mol Cell Biol* 2006, **7**:131-142.
41. Liotta LA: **Tumor invasion and metastases-role of the extracellular matrix: Rhoads Memorial Award lecture.** *Cancer Res* 1986, **46**:1-7.
42. Karpowicz P, Inoue T, Runciman S, Deveale B, Seaberg R, Gertsenstein M, Byers L, Yamanaka Y, Tondat S, Slevin J, Hitoshi S, Rossant J, van der Kooy D: **Adhesion is prerequisite, but alone insufficient, to elicit stem cell pluripotency.** *J Neurosci* 2007, **27**:5437-5447.
43. Karpowicz P, Willaime-Morawek S, Balenci L, DeVeale B, Inoue T, van der Kooy D: **E-Cadherin regulates neural stem cell self-renewal.** *J Neurosci* 2009, **29**:3885-3896.
44. Stremenova J, Krepela E, Mares V, Trim J, Dbaly V, Marek J, Vanickova Z, Lisa V, Yea C, Sedo A: **Expression and enzymatic activity of dipeptidyl peptidase-IV in human astrocytic tumours are associated with tumour grade.** *Int J Oncol* 2007, **31**:785-792.
45. Cheng GZ, Chan J, Wang Q, Zhang W, Sun CD, Wang LH: **Twist transcriptionally up-regulates AKT2 in breast cancer cells leading to increased migration, invasion, and resistance to paclitaxel.** *Cancer Res* 2007, **67**:1979-1987.
46. Cheng GZ, Zhang W, Wang LH: **Regulation of cancer cell survival, migration, and invasion by Twist: AKT2 comes to interplay.** *Cancer Res* 2008, **68**:957-960.
47. Kurrey NKKA, Bapat SA: **Snail and Slug are major determinants of ovarian cancer invasiveness at the transcription level.** *Gynecol Oncol* 2005, **97**:155-165.
48. Man TK, Chintagumpala M, Visvanathan J, Shen J, Perlaky L, Hicks J, Johnson M, Davino N, Murray J, Helman L, Meyer W, Triche T, Wong KK, Lau CC: **Expression profiles of osteosarcoma that can predict response to chemotherapy.** *Cancer Res* 2005, **65**:8142-8150.
49. Perez-Losada J, Sanchez-Martin M, Perez-Caro M, Perez-Mancera PA, Sanchez-Garcia I: **The radioresistance biological function of the SCF/klt signaling pathway is mediated by the zinc-finger transcription factor Slug.** *Oncogene* 2003, **22**:4205-4211.
50. Wang X, Ling MT, Guan XY, Tsao SW, Cheung HW, Lee DT, Wong YC: **Identification of a novel function of TWIST, a bHLH protein, in the development of acquired taxol resistance in human cancer cells.** *Oncogene* 2004, **23**:474-482.
51. Pure E: **The road to integrative cancer therapies: emergence of a tumor-associated fibroblast protease as a potential therapeutic target in cancer.** *Expert Opin Ther Targets* 2009, **13**:967-973.
52. Cheng GZ, Zhang WZ, Sun M, Wang Q, Coppola D, Mansour M, Xu LM, Costanzo C, Cheng JQ, Wang LH: **Twist is transcriptionally induced by activation of STAT3 and mediates STAT3 oncogenic function.** *J Biol Chem* 2008, **283**:14665-14673.
53. Mani SA, Guo W, Liao MJ, Eaton EN, Ayyanan A, Zhou AY, Brooks M, Reinhard F, Zhang CC, Shipitsin M, Campbell LL, Polyak K, Briskin C, Yang J, Weinberg RA: **The epithelial-mesenchymal transition generates cells with properties of stem cells.** *Cell* 2008, **133**:704-715.
54. Isenmann S, Arthur A, Zannettino AC, Turner JL, Shi S, Glackin CA, Gronthos S: **Twist family of basic helix-loop-helix transcription factors mediate human mesenchymal stem cell growth and commitment.** *Stem Cells* 2009, **27**:2457-2468.
55. Petit A, Sellers DL, Liebl DJ, Tessier-Lavigne M, Kennedy TE, Horner PJ: **Adult spinal cord progenitor cells are repelled by netrin-1 in the embryonic and injured adult spinal cord.** *Proc Natl Acad Sci USA* 2007, **104**:17837-17842.
56. **Bioconductor software.** [http://bioconductor.org].
57. **GoMiner.** [http://discover.nci.nih.gov/gominer/htgm.jsp].

doi:10.1186/1476-4598-9-194

**Cite this article as:** Mikheeva et al.: TWIST1 promotes invasion through mesenchymal change in human glioblastoma. *Molecular Cancer* 2010 **9**:194.

**Submit your next manuscript to BioMed Central and take full advantage of:**

- **Convenient online submission**
- **Thorough peer review**
- **No space constraints or color figure charges**
- **Immediate publication on acceptance**
- **Inclusion in PubMed, CAS, Scopus and Google Scholar**
- **Research which is freely available for redistribution**

Submit your manuscript at  
www.biomedcentral.com/submit

